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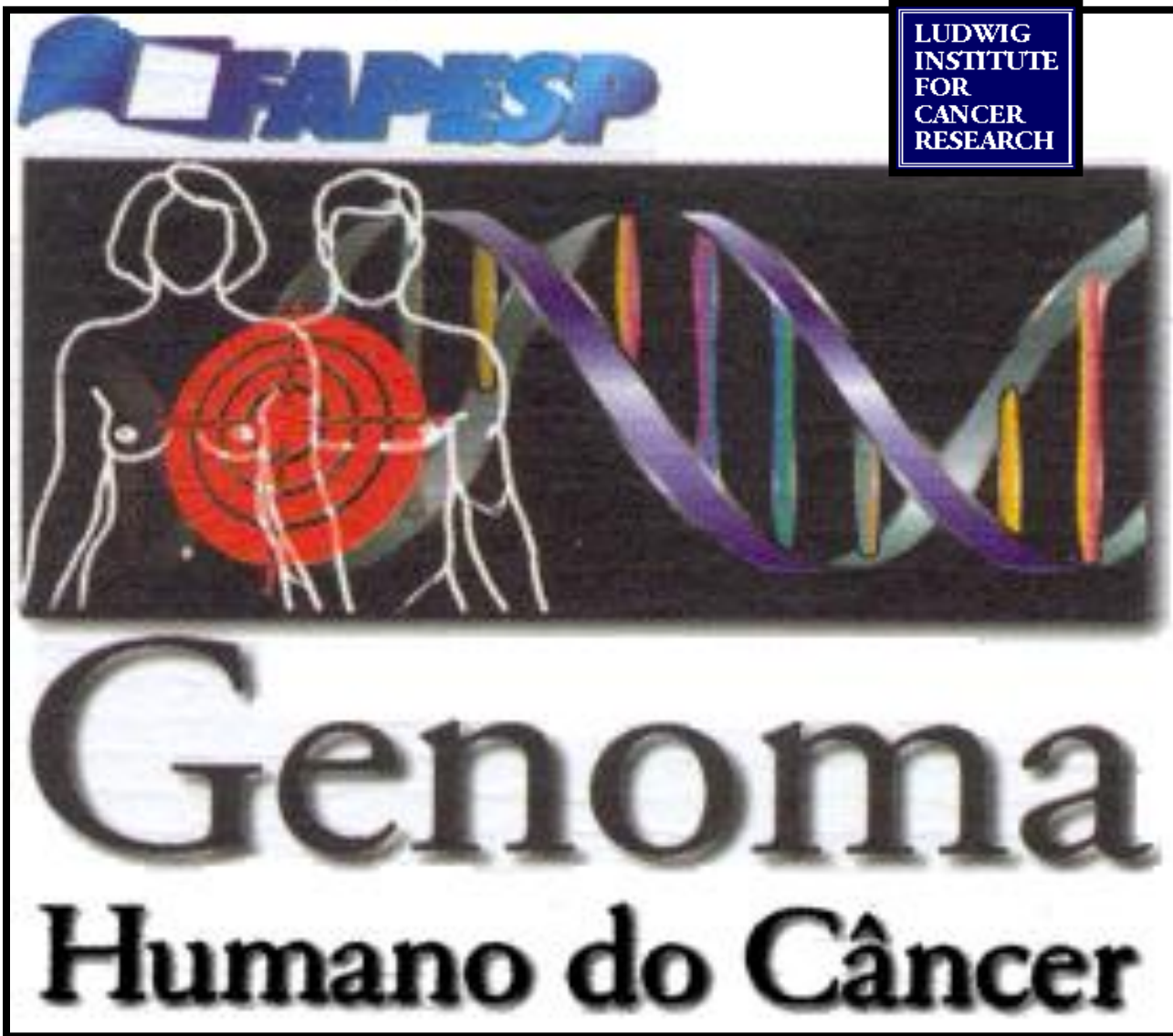
nature

Brazilian scientists team up for cancer genome project

[SÃO PAULO] Brazilian researchers have entered the competitive field of human genome sequencing with the signing of an agreement between the state funding agency of São Paulo (FAPESP) and the US-based Ludwig Institute for Cancer Research.

Each will contribute US\$5 million to a two-year Human Cancer Genome Project. According to FAPESP, the programme is "aimed at providing sequences from genes expressed in tumours that are important within the context of public health in the state of São Paulo".

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The FAPESP/LICR-Human Cancer Genome Project

RESEARCH NOTE

The Schistosome Genome Project: RNA Arbitrarily Primed- PCR Allows the Accelerated Generation of Expressed Sequence Tags

Emmanuel Dias Neto⁺, Richard Harrop^{**}, Rodrigo Corrêa-Oliveira, Sérgio DJ Pena^{*}, R Alan Wilson^{**}, Andrew JG Simpson^{***}

Centro de Pesquisas René Rachou - FIOCRUZ, Av. Augusto de Lima 1715, 30190-002 Belo Horizonte, MG Brasil ^{*}Departamento de Bioquímica e Imunologia, ICB-UFMG, Caixa Postal 486, 30161-970 Belo Horizonte, MG, Brasil ^{**}Department of Biology, University of York, Heslington, York, YO15DD, UK ^{***}Laboratório de Genética de Câncer, Instituto Ludwig de Pesquisas Contra o Câncer, R. Prof. Antônio Prudente 109/ 4^o andar, 01509-010 São Paulo, SP, Brasil

the databases before the project was initiated. The adoption of such a strategy proved to be extremely valuable in the analysis of abundant gene transcripts as well as in the discovery of new genes. However, there are some problems inherent to this method, such as the sequencing of vectors without insert and more importantly, the high level of redundancy. For example, of the 429 clones sequenced, 46 (10.7%) represented vectors without insert, and of 202 identified ESTs, homology with only 77 different genes was found, indicating a redundancy of 62% (Franco et al. 1995a *loc. cit.*). Whilst the former problem can be overcome, there is no simple way to solve the latter. These factors reduce the number of useful sequences obtained and thus increase the costs and the time required to tag all of the genes expressed by the parasite. In addition, to tag cDNAs derived from either rare, tissue or stage-specific mRNAs requires complex pre-processing of libraries, such as normalization, subtraction or differential hybridization (C Hoog 1991 *Nucl Acids Res* 19: 6123-6127).

With the aim of increasing the overall efficiency of EST generation, we have applied a technique of RNA arbitrarily primed PCR (RAP-PCR) to this process. RAP-PCR and its close relative "Differential Display", were first described by J Welsh et al. (1992 *Nucl Acids Res* 20: 7213-7218), and P

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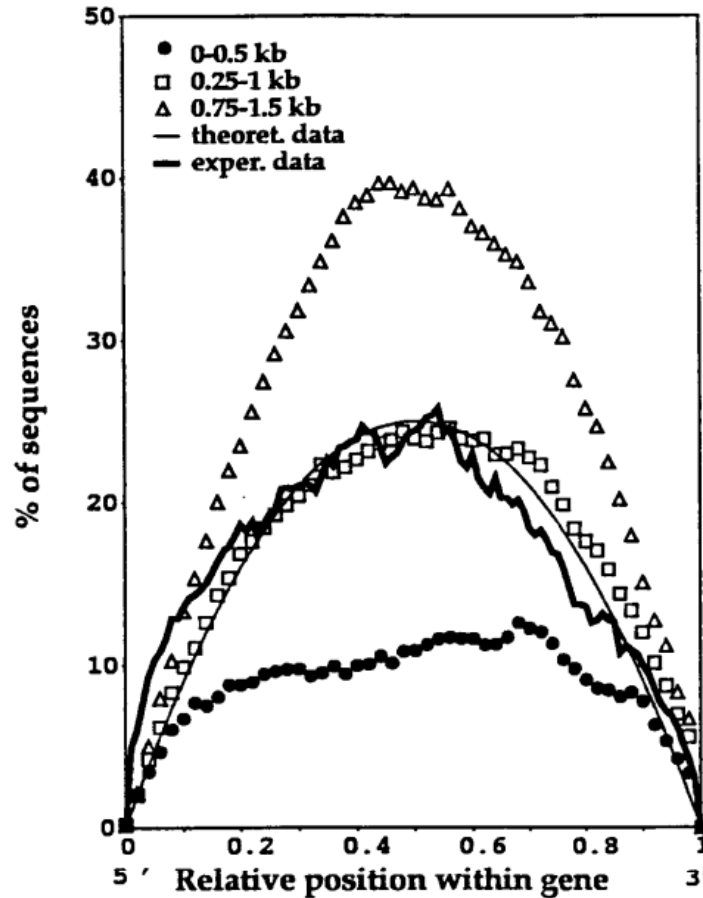
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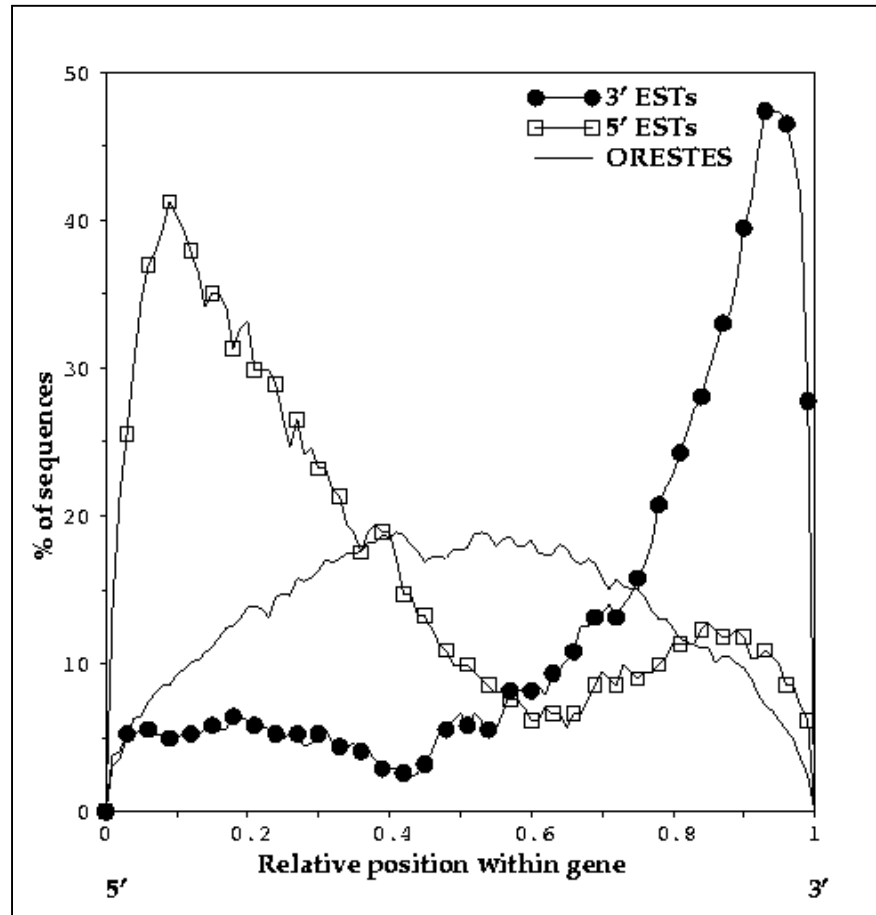


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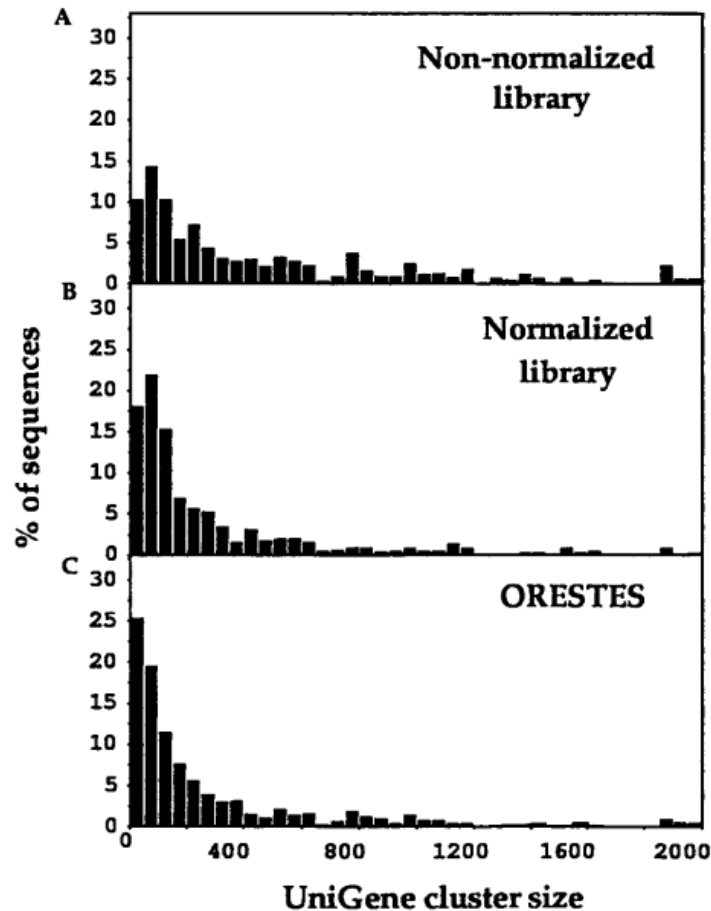


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Median cluster sizes
317, 138, and 125,

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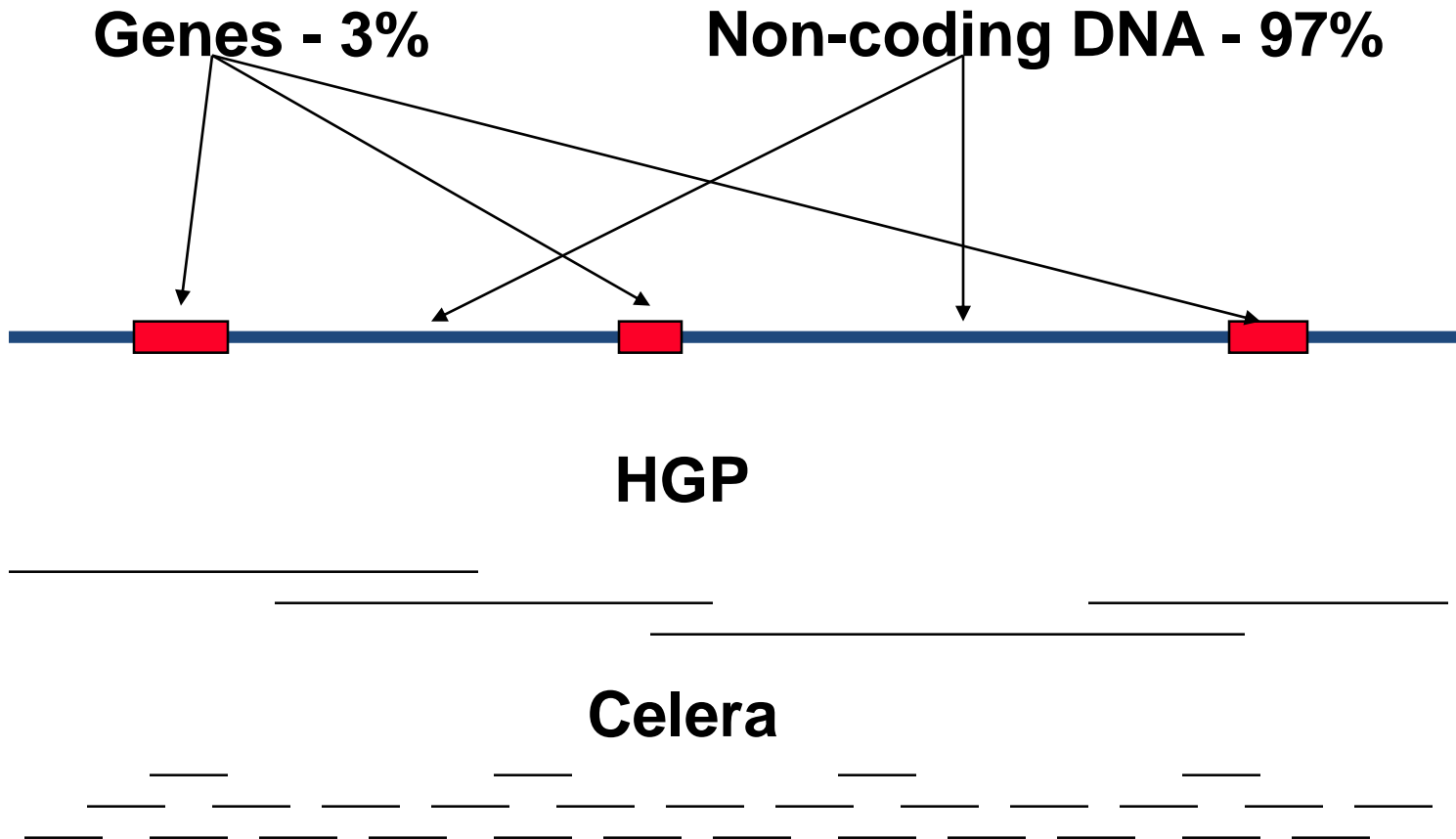
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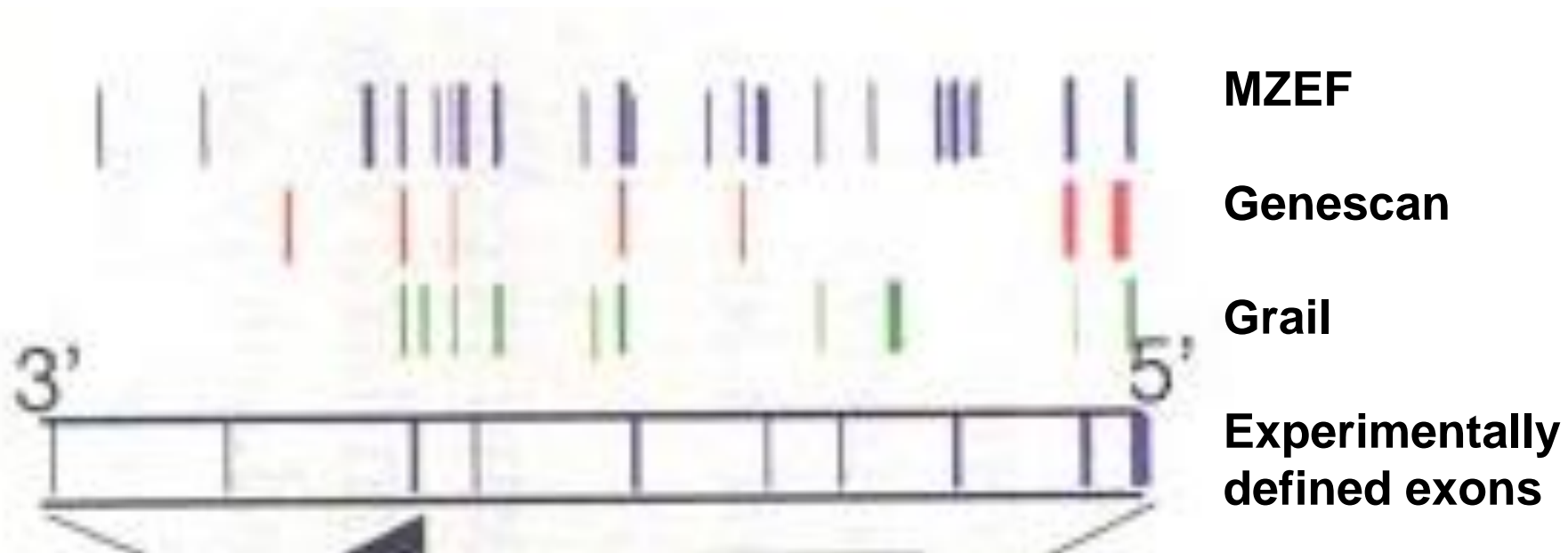


Human Genome Sequencing



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GCATTTAGAGGAAGTTTTGCTGATTCAGCGTGTGAATACGTGCCCAGAGGAGAGGAAGGGTCCGG
CTGACATTGAGTTATCTCTGC**AGGGCCTCTGATCCGGAAGCAGGTGTGGCCCGGGGCATAGACTT**
CCACCATGTGTCTGCTGTGCTCAACTTTGATCTTCCCCAACCCCTGAGGCCTACATCCATCGAG
CTGGCAGGTAGTAGTGTGACGGCCAGGCATCTGCATGGTAGGCACACTGAGGGACTTGGGGTGT
GCTGGACAGAGCCTGCGGGTTGGAGATGCAAGCTGCACTGTCTTCCCTTGC**AGGACAGCACGCGC**
TAACAACCCAGGCATAGTCTTAACCTTTGTGCTTCCCACGGAGCAGTTCACCTTAGGCAAGATTG
AGGAGCTTCTCAGTGGAGGTAAGAGCCTGGCTCTTGTGGTCCTGGGCCAGGGTCAGGCTTCTTCC
ACAATGCTTTAAACTCCATGATAATGATGACAGAGGTCACAACATAGTGTGACAGGCCACTTCC
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TGGGTGACGGGTCTTGGGCTTGGGGCCACACAGGCCACCCCCATCCACACACGGGGACAGATGG
GTCATCACTGTAAGAGGCCAGGTGCAGCTAACCTGCATGTTCCGCATCCAGGAAGGCGGTGGG
TCCCCTGCTGCTTTCCCCCAAGGGGGAGGTGCAGGAGGCCTCCAATGAAGACCCTATCCTAAGGC
CTCAGCCTGTGGGACCCTCGCTGCTTTCTTCTCCACAGAGAACAGGGGCCCCATTCTG**CTCCCCT**
ACCAGTTCGGATGGAGGAGATCGAGGGCTTCCGCTATCGCTGCAGGGTGAGCTGCTGTGGTGGG

Exon Prediction



Exon prediction in an A+T rich region of chromosome 21

- 250, 000 ORESTES
- 81,428 contigs
- 1,181 mapped to chromosome 22 (65.6% of 247 known genes*)
- 219 unannotated transcribed sequences of which 171 actually present in GenBank as cDNAs, 48 totally new

A = ORESTES predicted transcribed sequences
 B = Known genes *
 C = Genes predicted from other ESTs *

*

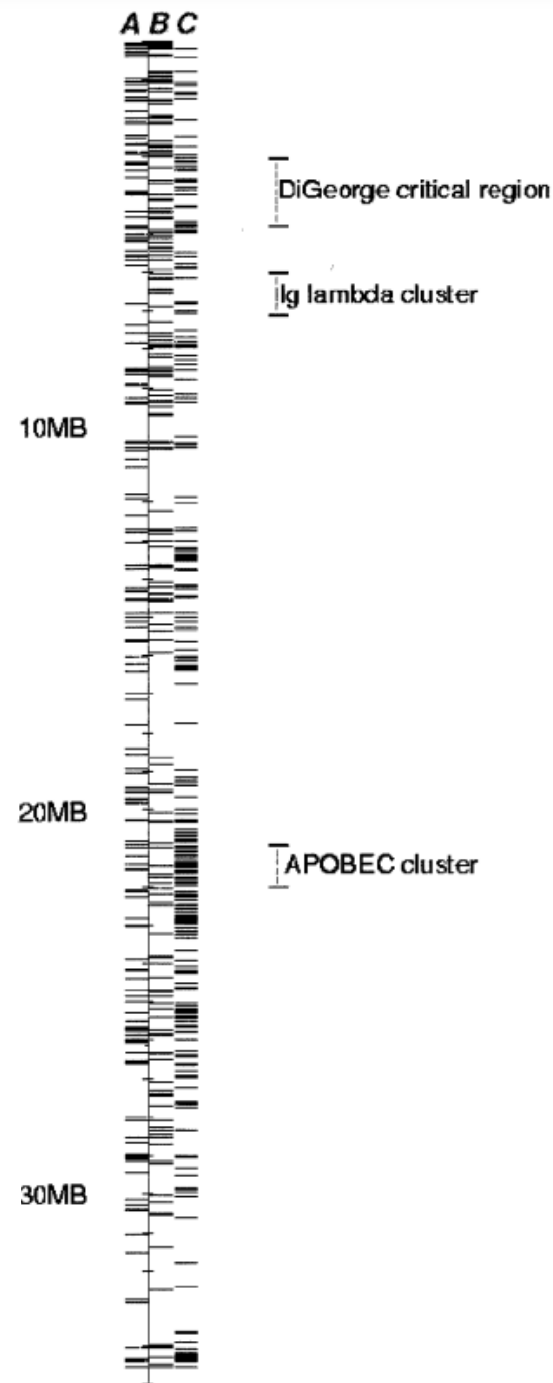
The DNA sequence of human chromosome 22

I. Dunham, N. Shimizu, B. A. Roe, S. Chisoe *et al.*†

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† A full list of authors appears at the end of this paper

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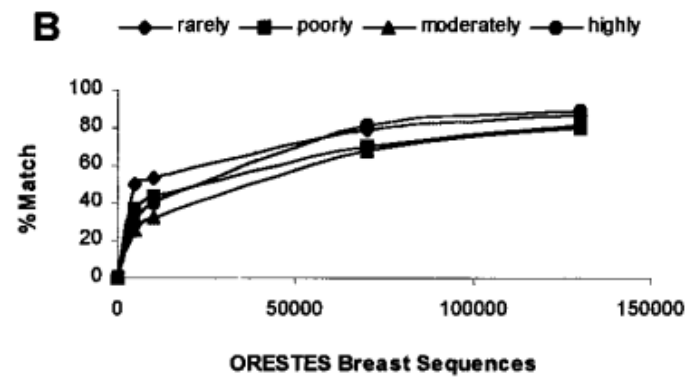
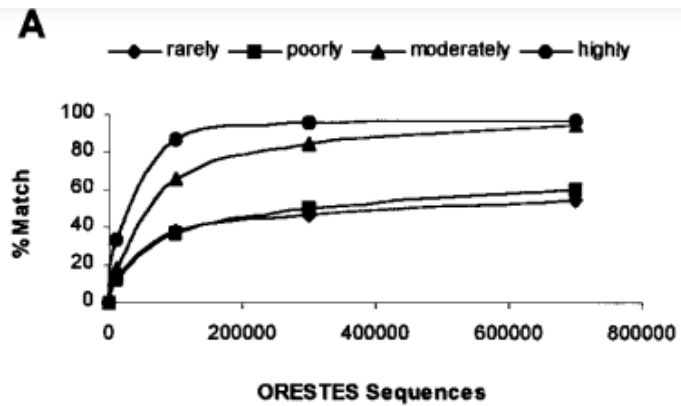
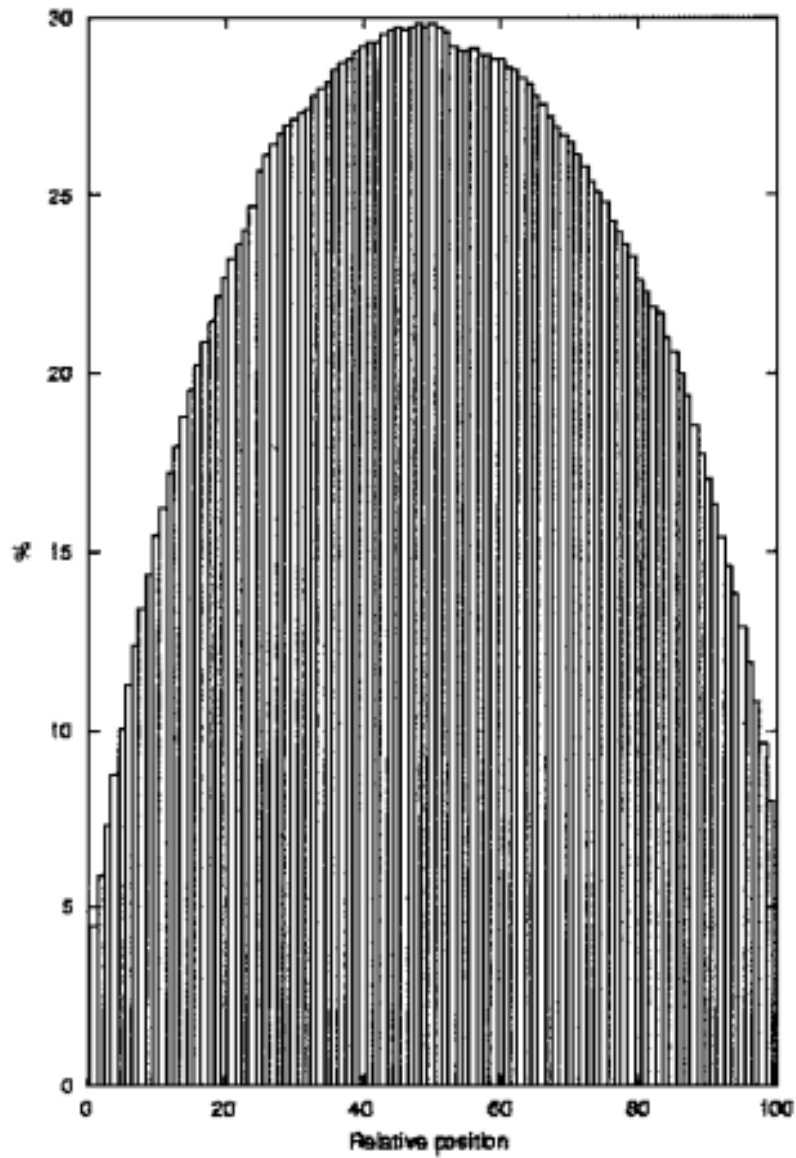
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In two large projects, extensive EST sequencing of human tumor tissues has been undertaken: the Cancer Genome Anatomy Project (CGAP) (3) and the Fundação de Amparo à Pesquisa do Estado de São Paulo/Ludwig Institute for Cancer Research–Human Cancer Genome Project (HCGP) (4, 5). CGAP, launched in 1997 by the National Cancer Institute, has used single-pass sequencing from the 5' and/or 3' extremities of cDNA clones for sequence generation (6). The HCGP project adopted an alternative EST-based strategy, termed ORESTES, which generates sequences biased toward the central coding regions of transcripts (7). The data gathered by these two projects are thus complementary and have been combined into an International Database of Cancer Gene Expression (5), available at <http://cgap.nci.nih.gov>. They also constitute the basis of the Human Cancer Index at TIGR (www.tigr.org/tdb/tgi/hcgi).

Comparative Study > Nat Genet. 2000 Jun;25(2):232-4. doi: 10.1038/76115.

Analysis of expressed sequence tags indicates 35,000 human genes

B Ewing ¹, P Green

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Initial sequencing and analysis of the human genome

International Human Genome Sequencing Consortium*

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The human genome holds an extraordinary trove of information about human development, physiology, medicine and evolution. Here we report the results of an international collaboration to produce and make freely available a draft sequence of the human genome. We also present an initial analysis of the data, describing some of the insights that can be gleaned from the sequence.

The rediscovery of Mendel's laws of heredity in the opening weeks of the 20th century^{1–3} sparked a scientific quest to understand the nature and content of genetic information that has propelled biology for the last hundred years. The scientific progress made falls naturally into four main phases, corresponding roughly to the four quarters of the century. The first established the cellular basis of heredity: the chromosomes. The second defined the molecular basis of heredity: the DNA double helix. The third unlocked the informational basis of heredity, with the discovery of the biological mechanism by which cells read the information contained in genes and with the invention of the recombinant DNA technologies of cloning and sequencing by which scientists can do the same.

The last quarter of a century has been marked by a relentless drive

coordinate regulation of the genes in the clusters.

- There appear to be about 30,000–40,000 protein-coding genes in the human genome—only about twice as many as in worm or fly. However, the genes are more complex, with more alternative splicing generating a larger number of protein products.
- The full set of proteins (the 'proteome') encoded by the human genome is more complex than those of invertebrates. This is due in part to the presence of vertebrate-specific protein domains and motifs (an estimated 7% of the total), but more to the fact that vertebrates appear to have arranged pre-existing components into a richer collection of domain architectures.
- Hundreds of human genes appear likely to have resulted from horizontal transfer from bacteria at some point in the vertebrate lineage. Diverse forms of gene transfer have been identified from trans-

Table 1. HCGP and CGAP transcript sequence generation and clustering

Form of gene representation	Number of sequences, clusters, or genes
ORESTES submitted to GenBank	823,121 sequences
CGAP EST submitted to GenBank	1,214,358 sequences
TOTAL EST submitted to GenBank	2,037,479 sequences
Total clusters	32,129 clusters
Total clusters with known genes	22,152 clusters
Clusters without known genes	9,977 clusters
Clusters without known genes but with coding potential	1,285 clusters
Estimated total genes based on HCGP and CGAP data	23,437 genes



A Transcript Finishing Initiative for Closing Gaps in the Human Transcriptome

The Ludwig–FAPESP Transcript Finishing Initiative,¹ Mari Cleide Sogayar,² and
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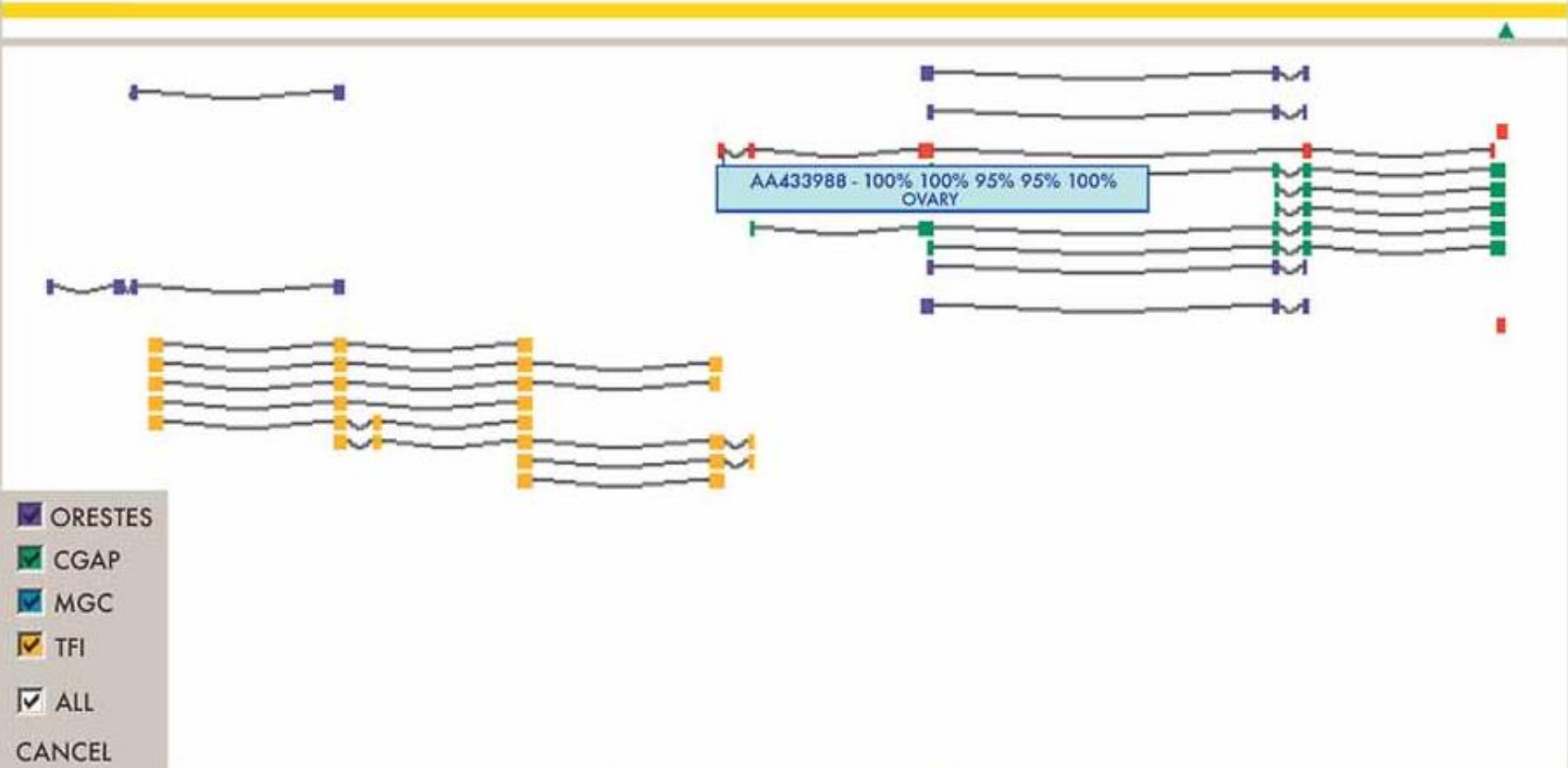
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TFU0171



We report the results of a transcript finishing initiative, undertaken for the purpose of identifying and characterizing novel human transcripts, in which RT-PCR was used to bridge gaps between paired EST clusters, mapped against the genomic sequence. Each pair of EST clusters selected for experimental validation was designated a transcript finishing unit (TFU). A total of 489 TFUs were selected for validation, and an overall efficiency of 43.1% was achieved. We generated a total of 59,975 bp of transcribed sequences organized into 432 exons, contributing to the definition of the structure of 211 human transcripts. The structure of several transcripts reported here was confirmed during the course of this project, through the generation of their corresponding full-length cDNA sequences. Nevertheless, for 21% of the validated TFUs, a full-length cDNA sequence is not yet available in public databases, and the structure of 69.2% of these TFUs was not correctly predicted by computer programs. The TF strategy provides a significant contribution to the definition of the complete catalog of human genes and transcripts, because it appears to be particularly useful for identification of low abundance transcripts expressed in a restricted set of tissues as well as for the delineation of gene boundaries and alternatively spliced isoforms.

Characterization of a cancer/testis (CT) antigen gene family capable of eliciting humoral response in cancer patients

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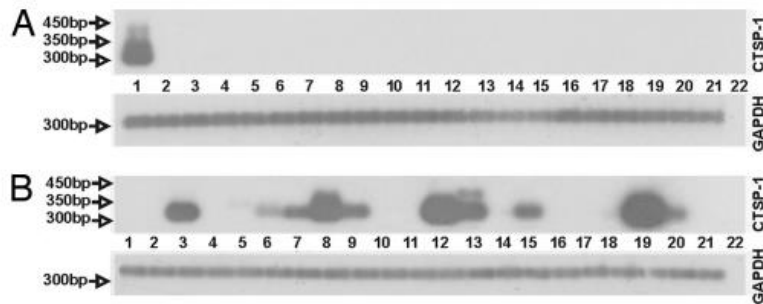


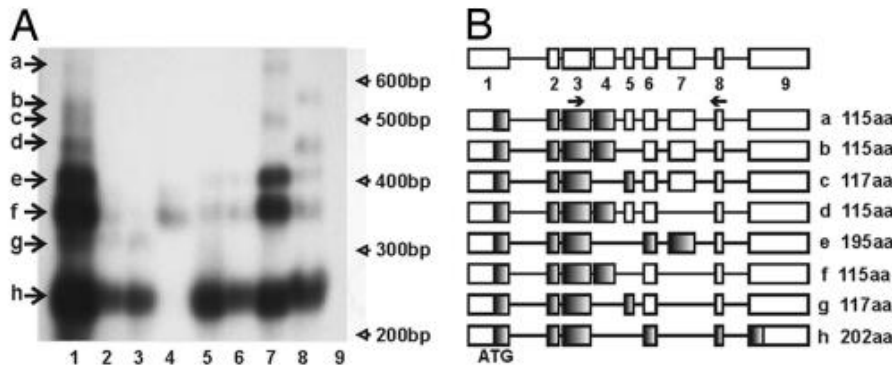
Fig. 4. CTSP-1 mRNA expression pattern in normal tissues and tumor cell lines. Southern blot of RT-PCR products amplified with CTSP-1-specific primers. (A) Normal cDNA samples used were as follows. Lanes: 1, testis; 2, lung; 3, prostate; 4, small intestine; 5, breast; 6, brain; 7, heart; 8, uterus; 9, bone marrow; 10, placenta; 11, colon; 12, fetal brain; 13, liver; 14, fetal liver; 15, thymus; 16, salivary gland; 17, spinal cord; 18, kidney; 19, spleen; 20, skeletal

Table 1. Frequency of mRNA expression of CTSP family members in tumor samples

Tumor	CTSP-1 (%)		CTSP-2 (%)		CTSP-4 (%)	
Breast	9/25	(36.0)	1/6	(16.5)	0/6	(0.0)
Colon	7/18	(39.0)	0/14	(0.0)	0/14	(0.0)
Esophagus	2/5	(40.0)	2/4	(50.0)	0/4	(0.0)
Glioblastoma	6/13	(46.0)	Not done		Not done	
Lung	8/14	(57.0)	2/11	(18.8)	0/11	(0.0)
Melanoma	10/17	(59.0)	1/9	(11.0)	0/9	(0.0)
Prostate	14/24	(58.0)	8/17	(47.0)	0/17	(0.0)
Stomach	4/9	(44.0)	1/3	(33.3)	0/3	(0.0)
Thyroid	7/24	(29.0)	0/6	(0.0)	0/6	(0.0)
Uterus	8/20	(40.0)	4/14	(28.5)	0/14	(0.0)
Total	75/169	(44.4)	19/84	(22.6)	0/84	(0.0)

Table 2. Frequency of anti-CTSP-1 antibodies in plasma samples from cancer patients

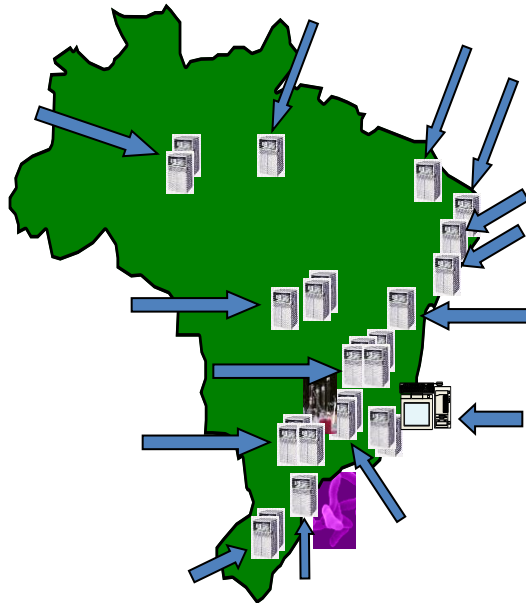
Tumor	Antibody response (%)	
Breast	3/18	(16.6)
Colon	0/20	(0.0)
Esophagus	0/4	(0.0)
Lung	1/13	(8.0)
Melanoma	1/22	(4.5)
Prostate	5/24	(20.8)
Stomach	1/8	(12.5)
Thyroid	2/10	(20.0)
Uterus	1/22	(4.5)
Total	14/141	(10.0)



The complete genome sequence of *Chromobacterium violaceum* reveals remarkable and exploitable bacterial adaptability

PNAS 100, 11660-11665, 2003

Brazilian National Genome Project Consortium*



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$$\underline{6G11+7C10+7A9+10G6+5A4}$$

MP RU IC JJ IL/II CB/EQ JJ CE RC IL/BF UV JE EZ

$$\underline{3H11+3E1+7B11+7E5+3C3+7B7+7G8+7C1}$$

$$\underline{8G8+7H3+2D9+5E12+10B10+5B8}$$

JJ/RC QR QR QR CB MR

$$\underline{2A11}$$

EZ

$$\underline{9A12+ 5B3+ 7A11+7A2+8A4+6D6+11A7+11A8+11A9+7B4+7C7+11D3+6D10+9H12+11A4}$$

QR JE MI CC PF MC MC IL CE AC BF CM QR QR AC

$$\underline{11A2 + 5F5+10H5+ 3H12+ 7C9} \quad 7F2$$

CC UT CC EP MP IC

$$\underline{11B8+9C12+11H5}$$

QV II EZ/EQ

$$\underline{4A4 +1G6 +9E10+ 9D10 + 7G2+ 11A5+ 7B1+ 7B12+ 7B5+ 11A10+ 1H9+ 4A9+ 1A1+ 3C12 +7C12}$$

QR UT/CE BG/MC EQ/CB JE RF EQ CB II IL QR QV PM MC AG

$$\underline{9A10+9H11+5G2+ 11A3}$$

QR JJ IU RF

$$\underline{1G4 + 7B9 + 7A12+ 3C11+ 8G12+ 9D11}$$

BG PM MR UV UV AC/BF

$$\underline{11G6+ 7C6}$$

EP/CM BG

$$\underline{1E1+ 2D3+ 7G7+ 2F10+ 6C10+ 7B9 + 8A2+}$$

AG/IU IU IC JJ AG JJ JJ

$$\underline{7A7+ 7H4+ 3D3+ 7B3+ 7B10 + 6C6}$$

UT QR QV QV RP RU/RP

$$\underline{9G12 + 7B2 +7C4 +6A8 +7A1 +6F5 +7A8}$$

PF PF JE QH CC IL RC

$$\underline{7A10+2E6+7A4+10F3+3A12}$$

QH QH UI EQ/CB IC

$$\underline{+8A3+ 9E9+5G3+8F1+2G4+ 8D10+ 7H2 + 6H3}$$

QV JJ JE JJ EZ MI QS QV/UT

$$\underline{2G12+1A3+19H11}$$

QS/QH UV JJ



Research Report

PCR-Assisted Contig Extension: Stepwise Strategy for Bacterial Genome Closure

BioTechniques 34:626-632 (March 2003)

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INTRODUCTION

Bacterial genome projects typically accrue most sequence data through high-throughput shotgun sequencing. The resulting shotgun draft is then converted into a complete representation of the organism's DNA through a more laborious completion phase. This phase is devoted mainly to the generation of additional sequence data to close gaps

that could, in principle, have been exploited for such a strategy. The majority of these have been utilized for the isolation of 5' flanking genomic regions of cDNA clones to clone insertion-tagged genes and to isolate the extremities of the inserts of large clones such as those in YACs (6–13). However, none of these methods have been previously evaluated as a strategy for bacterial genome closure. The majority

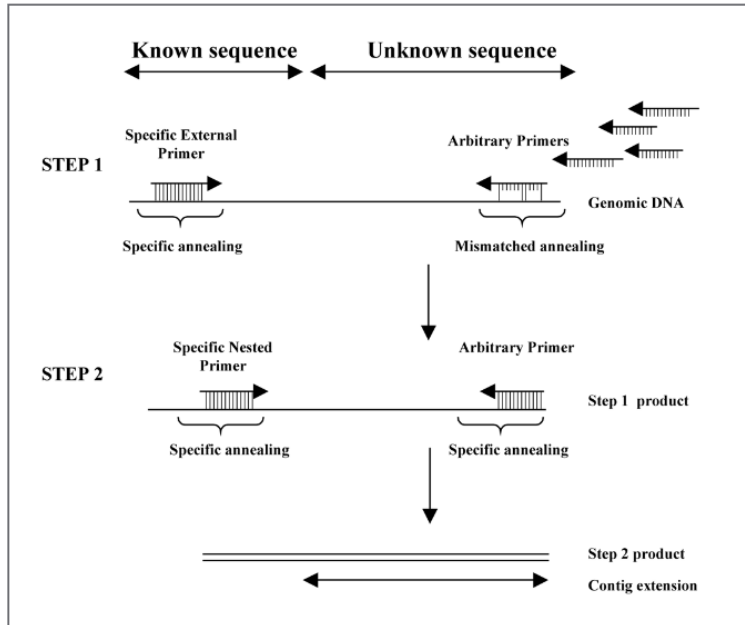


Figure 1. A schematic outline of the PACE reaction.

Table 1. Summary of Results Obtained with PACE in Extending *C. violaceum* Shotgun Contigs

Contig ^a	Amplification ^b		Single Products ^c		Sequences ^d		Specific Products ^e		Largest Single Product ^f (kb)
	step 1	step 2	step 1	step 2	step 1	step 2	step 1	step 2	
1	24	48	6	10	1	4	1	4	2.1
2	20	72	8	29	3	17	3	17	2.5
3	27	69	6	12	0	2	0	2	1.5
4	34	76	12	27	0	4	0	4	2.5
5	23	40	1	7	0	3	0	3	1.5
6	43	71	16	25	4	17	4	17	2
7	14	58	2	15	0	8	0	7	1.6
8	30	46	7	19	0	4	0	4	1.7
9	32	54	11	19	4	3	0	2	1.5
10	32	72	15	21	2	7	0	7	1.3
11	40	56	3	17	0	7	0	7	3
12	33	45	10	22	3	8	3	8	1.5
13	55	59	14	20	1	8	0	8	1.8
14	39	56	10	11	2	0	0	0	2.1
15	26	44	9	23	0	13	0	13	1.9
16	46	55	6	14	0	4	0	4	1.3
17	53	49	11	9	0	3	0	3	2.6
18	24	49	8	13	0	1	0	1	1.5
19	15	56	1	15	0	3	0	3	1.7
20	36	53	6	11	0	8	0	7	1.8
21	28	38	3	10	0	4	0	4	1.4
22	14	34	6	17	1	1	0	1	3
Total	688	1200	171	366	21	129	11	126	

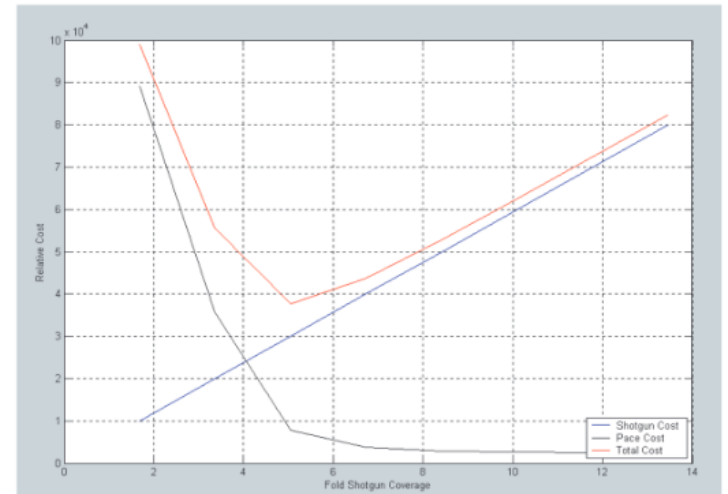
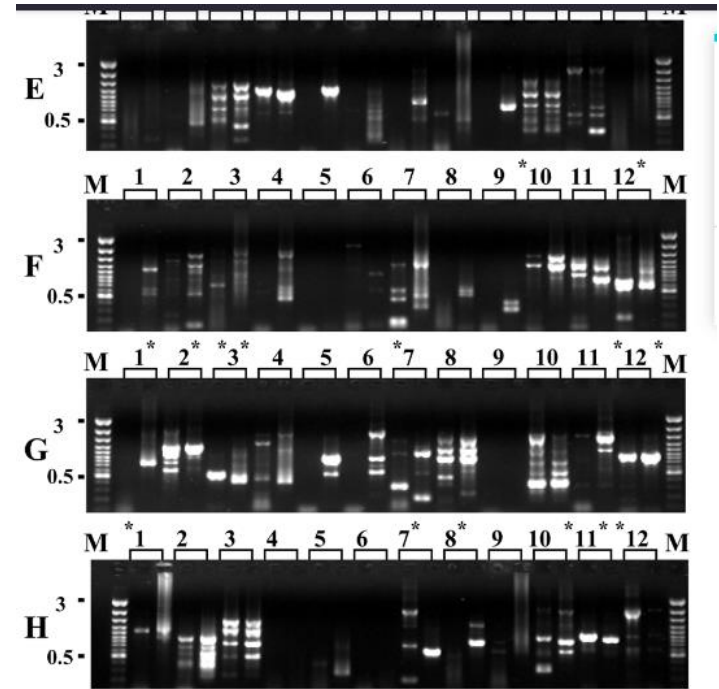


Figure 3. Relative overall cost of a complete 5-Mb genome sequence generated using PACE lowering different levels of shotgun coverage.

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The genome and transcriptomes of the anti-tumor agent *Clostridium novyi-NT*

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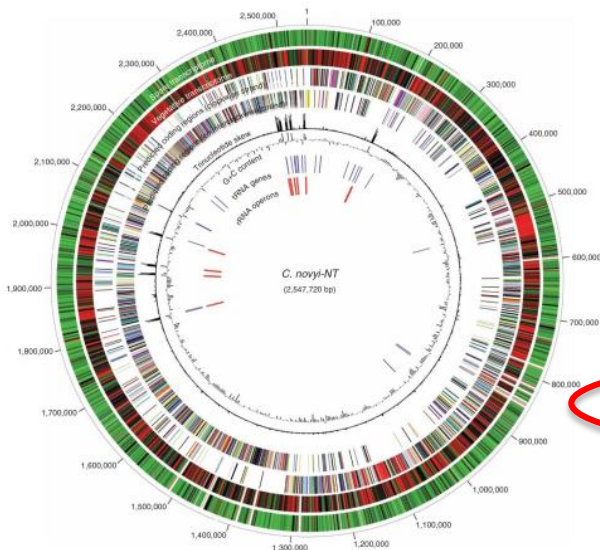


Figure 1. Circular representation of *C. novyi-NT* genome and transcriptomes. Predicted CDS are color coded based on functional classification. CDS in the transcriptome circles are also color coded, with red representing the highest mRNA abundance, green the lowest and white the intergenic regions. The G+C content and trinucleotide skew were determined using 2,000-nt windows with 1,000-nt incremental shifts.

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Brazil's biotech boom

Ten years ago, Brazilian bioscience was transformed by a bold initiative. Scientists and the government must develop and extend the progress that has resulted.

In May 1997, a pair of Brazilian scientists spent a weekend in the country discussing a bold idea. José Fernando Perez, the science director at the São Paulo Research Foundation (FAPESP), a state-funded agency and one of Brazil's leading research sponsors, had been looking for game-changing research initiatives. Biologist Fernando Reinach, one of his advisers, had a sufficiently adventurous plan: kick-start biotechnology research throughout Brazil by sequencing a genome.

For many risk-averse scientists in the old guard, who were acutely aware of how far the country lagged behind the rest of the world in biotechnology, this plan seemed overly ambitious. But the duo pushed ahead to build the capacity for genomics and bioinformatics that Brazil lacked, quickly organizing a team to conduct the project and then settling on a bacterium to sequence. FAPESP invested the equivalent of US\$12 million, largely dedicated to sequencers, computers and reagents, while the team brought together and trained researchers from a range of fields to develop a broad and long-lasting set of skills and knowledge.

On 13 July 2000 that effort paid off when the team, by then comprising more than 100 researchers in 35 Brazilian labs, published the genetic code for the citrus pathogen *Xylella fastidiosa* in an article featured on the cover of *Nature* (A. J. G. Simpson *et al.* *Nature* 406, 151–157; 2000). Ten years later, the fruits of that project keep coming.

Before its *Xylella* paper had even come out, for example, the network was busy sequencing another citrus pathogen while taking its first stab at the complex sugarcane genome and contributing to the

Brazilian biotechnology has matured to the point at which its scientists are players on the international stage. And FAPESP is still promoting big ideas, including a new programme to pump money into a broad portfolio of bioenergy research even as the Ministry of Science and Technology constructs a bioethanol research centre; both initiatives seek to build on Brazil's lead in this field. FAPESP is also working to overcome one of the biggest impediments to progress — a lack of doctoral researchers — by encouraging scientists to fill the gaps with young stars from the United States and Europe, part of a broader effort to internationalize Brazilian science.

All of this is good, but more efforts are needed in the same vein — more attitude, more risk-taking and more entrepreneurialism that puts public science into private practice, an area in which Brazil continues to lag. Universities and funding agencies must continue to advance technology-transfer programmes, and the government must streamline regulations that slow even simple activities such as purchasing scientific equipment from abroad. But if there is anything holding Brazil back, it is the same unjustified fear of failure that the country overcame ten years ago with *Xylella*. Although institutions can promote, fund and reward bold thinking, it is worth noting that *Xylella* was not simply a bricks-and-mortar research centre run by a foundation, but a science project. Ultimately the task of promoting Brazilian biotechnology comes down to the science, and it will be up to individual scientists to accept the challenge and expand their research horizons.



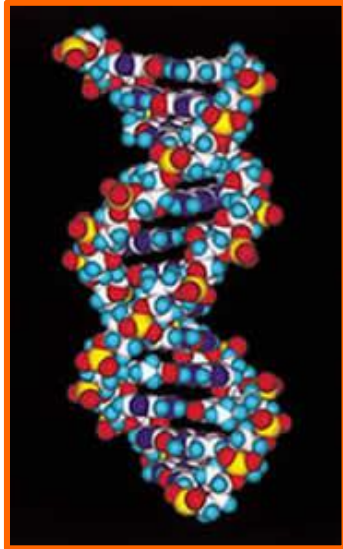
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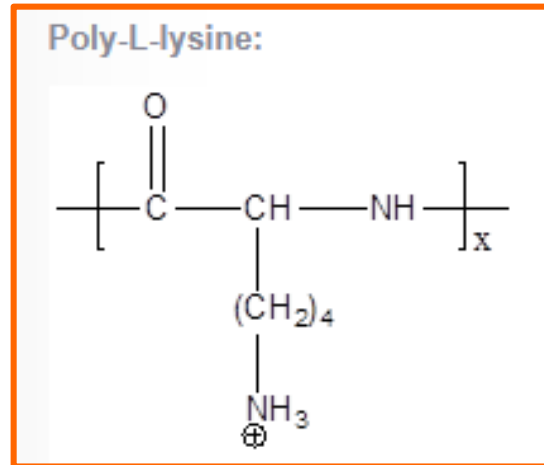
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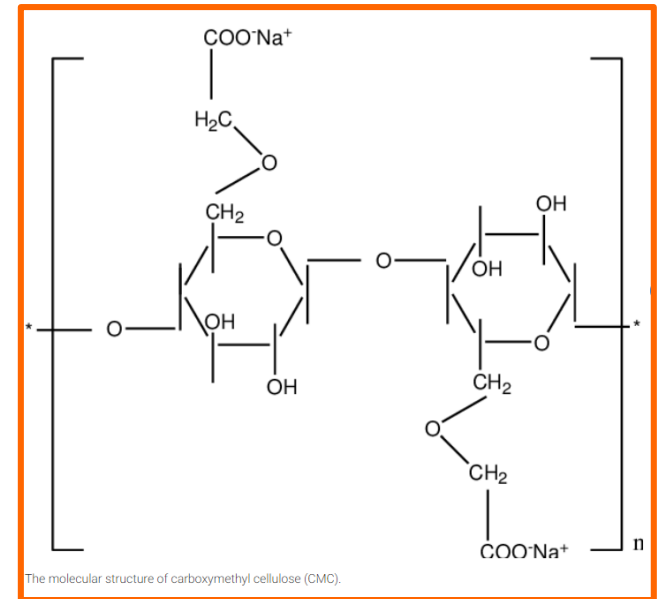
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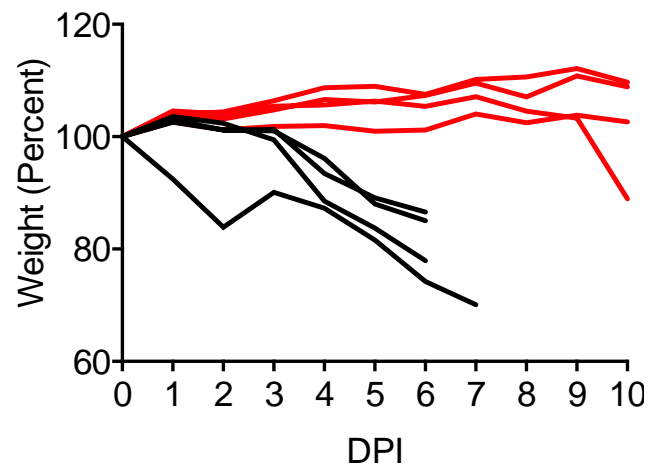
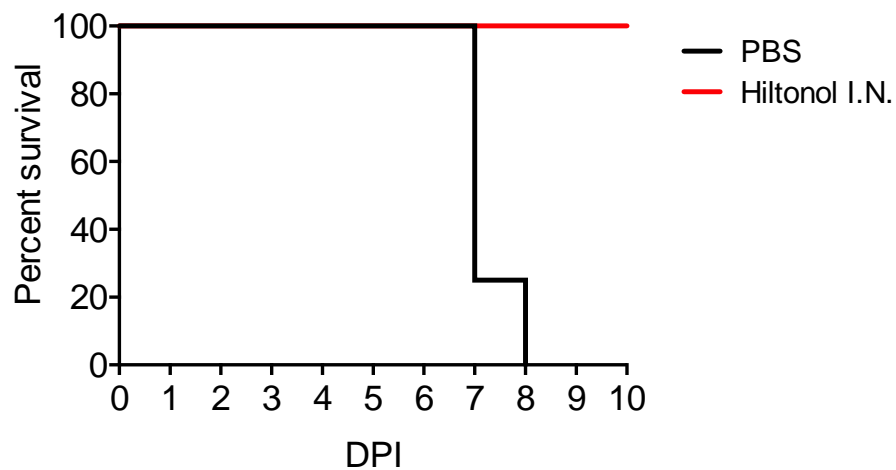
+ carboxymethylcellulose

Poly-ICLC is a stabilized form of double stranded RNA that mimics a viral genome and acts as a potent and rapid activator of the immune response to be used either alone or with an antigen to direct the immune response to tumors or infectious agents being developed by **Oncovir**.

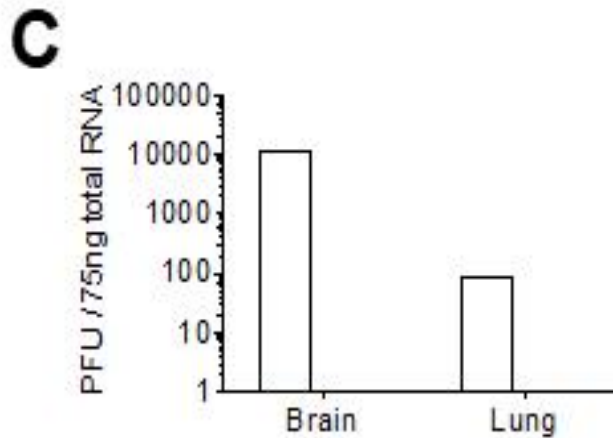
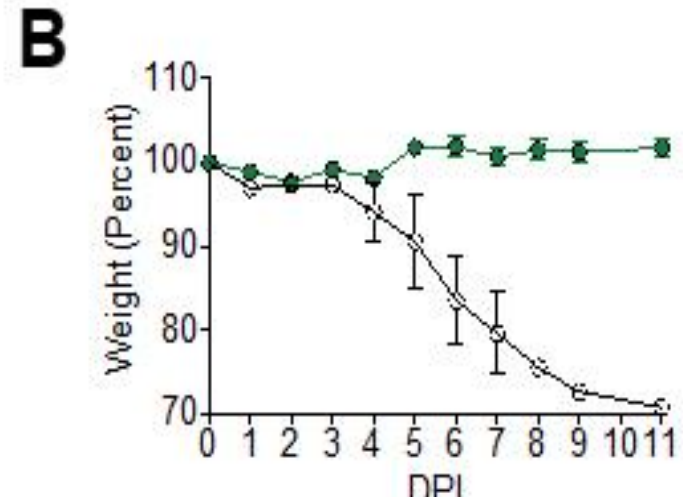
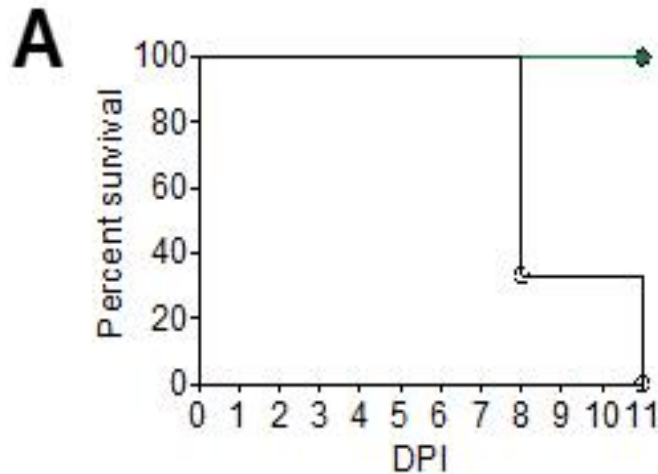
Supplied as a 1.8mg/ml sterile solution in saline in vials containing 1.2ml

Proteção contra COVID

Mice: 50ug (28uL) Hiltonol i.n. (days -4 and -3)



Vacina contra COVID



nature communications



Article

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Promotion of neutralizing antibody-independent immunity to wild-type and SARS-CoV-2 variants of concern using an RBD-Nucleocapsid fusion protein

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Check for updates

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FAPESP

Genoma



Organization for
Nucleotide
Sequencing and
Analysis

The Virtual Genomics Institute